



# ELEGANCE

## URINARY GROWTH HORMONE ELISA KIT

REF 40 GHU96  
Σ 96



### WARRANTY

The manufacturer makes no express warranty other than the diagnostic kit will measure the designated analyte when used in accordance with the manufacturer's printed instructions. The use of the diagnostic kit for any other purpose is outside the intended use of this product and is done at the user's own risk.

The manufacturer disclaims any and all implied warranties of merchantability, fitness for use or implied utility for any other purposes. Any and all damages for failure of the diagnostic kit to perform according to its instructions are limited to the replacement value of the kit.

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### INTENDED USE

The *ELEGANCE* UGH ELISA has been designed for the quantitative *in vitro* diagnostic measurement of urinary growth hormone (UGH) in urine.

### PRINCIPLES OF THE ELEGANCE ELISA

The *ELEGANCE* ELISA is an enzyme-linked immunoassay. The sample reacts with anti-human GH antibodies bound to the microwell, and further reacts with the biotinylated antibodies.

The microwells are washed to remove any unbound material. Streptavidin-peroxidase (Amplification Reagent) is added and binds to the biotinylated antibody at many sites. After washing, the substrate solution reacts with any bound peroxidase to produce colour in direct proportion with the amount of sample antigen, which can be calculated from the calibrator curve.

### ELEGANCE REAGENTS PROVIDED, STABILITY AND STORAGE

Kit size - 96 tests. The kit and all its components, unopened or opened, should be stored at 2-8°C until the listed expiry dates.

### GH: Coated Microwells

**96 wells REF # GHA96**  
Frame containing microwells coated with anti-GH antibody. Ready to use.

### GH: Antibody Reagent

**1 vial REF # GHB96**  
10 mL biotinylated anti-GH antibody in a buffered solution containing bovine serum albumin, non-immune animal sera and a blue dye. Contains sodium azide, 0.2% w/v and thiomersal, 0.01% w/v. Ready to use.

### GH: Amplification Reagent

**1 vial REF # GHP96**  
10 mL streptavidin-peroxidase (streptavidin from *S. avidinii*) in a buffered solution containing bovine serum albumin and a violet dye. Contains Bronidox L, 0.2% v/v and thiomersal, 0.02% w/v. Ready to use.

### Wash Concentrate

**1 vial REF # EWC96**  
50 mL of a 15 x concentrated wash solution. Contains thiomersal, 0.09% w/v. To be diluted before use.

### Substrate Solution

**TMB H**  
**1 vial REF # ETMB96**  
10 mL 3,3',5,5'-tetramethylbenzidine and hydrogen peroxidase in a stabilising solution. Ready to use.

### UGH: Calibrators

**6 vials REF # HGHUS1-6**  
1 mL each in 1% BSA PBS buffer. Contains sodium azide, 0.1% w/v. Lyophilized.

### PRECAUTIONS AND WARNINGS TO USERS

Handling of specimens and kit components, their use, storage and disposal should be in accordance with any local or national laboratory safety procedures or regulations.

### Specimens and Calibrators

The source material of the calibrators has been tested by an approved accredited method for the presence of hepatitis B surface antigen, antibody to hepatitis C and antibody to HIV - 1/2 (AIDS) and has been found to be non-reactive for all. However, it is recommended that all samples be handled as if capable of transmitting infectious disease.

### Preservatives

The kit contains sodium azide, thiomersal and Bronidox L as preservatives. As reagents contain potentially toxic preservatives, care should be taken in handling, to avoid ingestion or skin contact. Sodium azide may react with lead and copper plumbing to form potentially explosive azides.

### Substrate

Avoid any skin contact.

### SPECIMEN COLLECTION AND HANDLING

No special patient preparation is required. Specimens are urine and collected in a manner appropriate for laboratory testing. Specimens can be stored at 2-8°C for up to 48 hours. Specimens held for longer should be stored at or below -20°C. Specimens should not be frozen and thawed repeatedly. Thawed specimens should be checked for flocculent matter and mixed by inversion just prior to testing.

### MATERIALS AND EQUIPMENT REQUIRED BUT NOT PROVIDED

- \* Distilled or deionised water
- \* 2M HCl
- \* Precision pipettes
- \* Repeating pipette
- \* 1L measuring cylinder
- \* Absorbent tissue (lint-free)
- \* Timer
- \* Vortex mixer
- \* Microtitre plate shaker
- \* Microtitre plate washer
- \* Microplate reader system.

### PROCEDURAL NOTES

Bring all reagents and specimens to room temperature (20-25°C) and mix by gentle inversion prior to use. Duplicates are recommended. Contamination of reagents will lead to poor performance. A calibrator curve should be run with each assay. Specimens suspected of having concentrations above the top calibrator should be diluted in zero calibrator before assay. All assay steps should be performed without interruption, but if the wells cannot be filled with Amplification Reagent or substrate solution immediately after washing, then the microwells may be left upside down on absorbent lint-free tissue for a maximum of 15 minutes.

Reagents are matched in each kit and therefore reagents from different lot numbers should not be mixed.

The photometer and all pipettes used should be calibrated appropriately before use.

### Washing

The efficiency of the wash step is vital for good precision. Microwells are washed using an automatic plate washer. Avoid overflows from one well to another.

### Quality Control

Control specimens should be run in every assay to ensure correct procedure. Control values should lie within laboratory ranges before assay is approved.

### ASSAY PROCEDURE

#### Preparation of Reagents Wash Solution

Dilute the wash concentrate 1 in 15 with deionised water. The wash solution can be stored at room temperature (20-25°C) for 12 weeks.

#### Calibrators

To reconstitute the lyophilized calibrators, add the volume of deionized water indicated on each vial label. Allow the vials to sit undisturbed until completely dissolved (at least 30 minutes) and then mix by gentle inversion. Exact concentrations determined lot-to-lot are stated on a separate label inside the kit. After reconstitution, the calibrators should be stored at -20°C for up to 4 weeks.

#### Protocol

1. Assemble the microwells in the frame according to the number of tests required. Bag and return unused wells to 2-8°C.
2. Pipette 200 µL of sample (calibrator, control, specimen) in duplicate into the appropriate wells. Time taken to dispense the samples should not exceed 20 minutes.

3. Cover microwells with lid and incubate for 90 minutes on a plate shaker at room temperature (20-25°C).

4. After incubation, wash the microwells. Aspirate the liquid and rinse each well 4 times with 300 µL wash solution. After the final wash, invert the microwells and tap firmly on absorbent tissue to remove any remaining wash solution. Ensure that no air bubbles remain in the wells.

5. Pipette 100 µL of GH Antibody Reagent (blue) into all wells.

6. Cover microwells with lid and incubate for 90 minutes on a plate shaker at room temperature (20-25°C).

7. After incubation, repeat wash step.

8. Pipette 100 µL of GH Amplification Reagent (violet) into all wells.

9. Cover microwells with lid and incubate for 30 minutes on a plate shaker at room temperature (20-25°C).

10. After incubation, repeat wash step.

11. Pipette 100 µL of Substrate Solution into all wells. Timing of the incubation step is measured from the addition of Substrate Solution to the first well.

12. Cover microwells with lid and incubate for 15 minutes stationary at room temperature (20-25°C).

13. Pipette 50 µL of 2M HCl into all wells in the same timed sequence as for Substrate Solution addition.

14. Carry out an end-point reading at 450 nm and process data as described in the microplate reader user's manual. This reading step should be carried out within 30 minutes of stopping the reaction.

### CALCULATION OF

#### RESULTS

Calculation of results can be carried out manually if there is no automatic data reduction. Determine the OD for each well. Plot the calibrator curve using log-log graph paper with concentration of calibrators on the x-axis and OD on the y-axis. The curve may be drawn point-to-point or a curve-fitting routine, such as spline interpolation, may be used. Interpolate the sample values from OD measured from this calibrator curve. Record the value for each sample in pg/mL UGH.

The range of the *ELEGANCE* UGH ELISA is from 0 to approx. 200 pg/mL, but the maximum concentration that can be reported is limited by the linear performance characteristics of the photometer used. If the OD value of the highest calibrator is above the range of the photometer, then this calibrator must be omitted from the plot of the calibrator curve.

#### MODEL CALCULATIONS

Endpoint Data		
ID	Ave OD	UGH (pg/mL)
0	0.152	
5	0.223	
10	0.332	
50	1.028	
100	1.827	
200	3.193	
Sample 1	0.440	15.85
Sample 2	1.027	48.20
Sample 3	1.891	103.43

#### CALIBRATION

The calibrators supplied in this kit are calibrated and labelled in pg/mL, referenced to the WHO 1988 1st IS 80/505.

#### Accuracy

Recovery was calculated by assaying before and after addition of exogenous analyte.

Sample	UGH (pg/mL) Observed	UGH (pg/mL) Expected	% Recovery
1	36.4	38.3	95.0
2	64.8	65.0	99.7
3	123.5	120.8	102.2

#### Dilution

A sample was diluted in zero calibrator, assayed and recovery calculated.

Sample	UGH (pg/mL) Observed	UGH (pg/mL) Expected	% Recovery
Neat	114.7		
1/2	54.8	57.3	95.6
1/4	26.4	28.7	92.0
1/8	12.7	14.3	88.8
1/16	7.1	7.2	98.6

#### Sensitivity

Measurement was carried out 5 times or more using 0 pg/mL and 5 pg/mL urinary GH standards. Mean of OD values and SD were calculated.

“Mean for 0 pg/mL + 2SD” was smaller than

“Mean for 5 pg/mL standard - 2SD”.

#### Interference

No interference with analyte recovery was observed for concentrations of haemoglobin up to 500 mg/dL, Free bilirubin up to 20 mL/dL, Conjugated bilirubin up to 20 mL/dL and Chyle up to 2,000 degrees (turbidity).

#### ORDERING INFORMATION

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#### TECHNICAL SERVICE

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PART No.: *EKBUGH Ed.7*

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#### EXPECTED VALUES

It is recommended that each laboratory establish its own reference range based on a representative sample population.

#### PERFORMANCE CHARACTERISTICS

##### Intra-assay Precision

Sample	n	Mean ± 2SD (pg/mL)	%CV
1	10	14.41 ± 0.70	2.4
2	10	42.46 ± 2.22	2.6
3	10	98.69 ± 6.05	3.1

##### Inter-assay Precision

Sample	n *	Mean ± 2SD (pg/mL)	%CV
1	16	15.68 ± 4.24	13.5
2	16	40.08 ± 7.94	9.9
3	16	73.39 ± 10.98	7.5

\* duplicate

#### Specificity

The OD for controls were in the range of 80-120% of its known concentration.